# **Accurate prediction of functional enhancer-promoter** interactions using epigenomic data

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## Motivation

- Transcriptional enhancers control how genes are expressed in specific cell types.
- Enhancer disruption and misregulation are implicated as disease-driving mechanisms.
- Modalities that specifically target enhancers that control disease-associated genes are being pursued to develop new drugs for a range of indications.
- However, it remains a major challenge to link functional enhancers to their target genes.

Results **EPIC outperforms ABC model in predicting enhancer**promoter pairs (holdout test data)





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Enhancer-promoter interaction characterization (EPIC) is a machine learning model for predicting functional enhancerpromoter (E-P) pairs.

Approach



#### **Basic features**

- HiChIP.AnchorSize: AnchorSize = 5kb, 10kb, 15kb, or 20kb (n=4)
- Assay.Position.WindowSize, where Assay=ATAC, H3K27ac, H3K4me1, H3K4me3, EP300, CTCF, or Input ChIP; Position = Enh or TSS; WindowSize = 300bp, 500bp, 1kb, 2kb, or 4kb (n=7\*2\*5=70)
- Genomic distance (n=1)

#### Feature engineering APMI = (ATAC.Enh.1kb \* EP300.Enh.1kb \* H3K4me1.Enh.4kb)<sup>1/3</sup> \* HiChIP.5kb



Model	AUPR	AUROC
EPIC-full	0.613	0.918
EPIC-basic	0.551	0.912
ABC	0.451	0.885

- The area under receiver operating characteristic (AUROC) curve of EPIC-full is significantly higher than that of ABC (p = 1.6e-10) (DeLong, et al., 1988).
- The AUROC of EPIC-full is significantly higher than that of EPIC-basic (p = 0.01), demonstrating the value of feature engineering.

Engineered features rank highest in feature importance.



Based on APMI, we engineered a new set of features for quantifying the relative contribution of an enhancer **e** to a gene **g** from the gene perspective or enhancer perspective:

$$fracGene_{eg} = \frac{APMI_{eg}}{\sum_{i} APMI_{ig}}$$

where j indexes all the enhancers connected to gene g.

**APMI**<sub>ea</sub>  $fracEnh_{eg} = \frac{1}{\sum_{k} APMI_{ek}}$ 

where k indexes all the genes connected to enhancer e. In addition, we combined these features to form new features.

 $fracGmE_{e,q} = fracGene_{e,q} * fracEnh_{e,q}$  $fracGpE_{e,g} = fracGene_{e,g} + fracEnh_{e,g}$ apmiGene<sub>e,g</sub> = fracGene<sub>e,g</sub> \* APMI<sub>e,g</sub> apmiEnh<sub>e,g</sub> = fracEnh<sub>e,g</sub> \* APMI<sub>e,g</sub> Enh1 - GeneA fracGene  $\frac{5}{1+1+5} = 0.71$ apmiGmE<sub>eg</sub> = fracGmE<sub>eg</sub> \* APMI<sub>eg</sub>  $\frac{5}{1+5} = 0.83$ fracEnh  $apmiGpE_{eg} = fracGpE_{eg} * APMI_{eg}$ 

### EPIC outperforms ABC model in linking GWAS loci to causal genes in a new cell type

- We generated epigenomic data in human primary hepatocytes and discovered about 30,000 E-P interactions using EPIC.
- Evaluate prediction of causal genes for liver-related GWAS loci using a curated set of "gold standard" locus-gene pairs (Mountjoy, et al., 2021)
  - Positive: GWAS locus-gene pairs in the gold standard set
  - Negative: GWAS loci connecting to other genes within 500kb





Donor IV

OTC c.-106C>A (Allele ID 480410, late-onset OTC deficiency) - pathogenic (dbSNP: rs749748052). leading to decreased OTC mRNA. Variant associated with 10-25% of normal OTC activity

#### Conclusions