# Linking GWAS risk variants to disease genes by epigenomic mapping and prediction of functional enhancer-promoter interactions ©2022

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## Motivation

- □ Majority of GWAS loci are noncoding, strongly enriched in gene regulatory elements such as enhancers.
- Identifying the genes regulated by these enhancers promises to reveal disease mechanisms.
- However, it remains a major challenge to link functional enhancers to their target genes.

# Results

**EPIC outperforms ABC model in predicting** enhancer-promoter pairs (holdout test data)

ABC = (ATAC.Enh.500bp \* H3K27ac.Enh.500bp)<sup>1/2</sup> \* HiC.5kb (Fulco et al., 2019)



**EPIC outperforms ABC model in linking GWAS** loci to causal genes in a new cell type

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- We generated epigenomic data in human primary hepatocytes and discovered about 30,000 E-P interactions using EPIC.
- Evaluate prediction of causal genes for liver-related GWAS loci using a curated set of "gold standard" locusgene pairs (Mountjoy, et al., 2021)
  - > Positive: GWAS locus-gene pairs in the gold standard set

## Approach

Enhancer-promoter interaction characterization (EPIC) is a machine learning model for predicting functional enhancer-promoter (E-P) pairs.





- The area under receiver operating characteristic (AUROC) curve of EPIC-full is significantly higher than that of ABC (p = 1.6e-10) (DeLong, et al., 1988).
- The AUROC of EPIC-full is significantly higher than that of EPIC-basic (p = 0.01), demonstrating the value of feature engineering.

Engineered features rank highest in

feature

apmiGmE -				
fracGmE =				
apmiGpE -				
fracGpE -				

> Negative: GWAS loci connecting to other genes within 500kb



Enhancers for CYP7A1 overlap with fine mapped variants across 15 lead SNPs representing associations from 32 GWAS studies of cholesterol (total, LDL, HDL), triglyceride levels, cholelithiasis, and cholestasis. CYP7A1 is a well characterized enzyme regulator of bile acid and cholesterol homeostasis and these enhancers have been experimentally validated (Wang, et al., 2018).

• HiChIP.AnchorSize: AnchorSize = 5kb, 10kb, 15kb, or 20kb (n=4) • Assay.Position.WindowSize, where Assay=ATAC, H3K27ac, H3K4me1, H3K4me3, EP300, CTCF, or Input ChIP; Position = Enh or TSS; WindowSize = 300bp, 500bp, 1kb, 2kb, or 4kb (n=7\*2\*5=70) • Genomic distance (n=1)

#### Feature engineering

APMI =  $(ATAC.Enh.1kb * EP300.Enh.1kb * H3K4me1.Enh.4kb)^{1/3} *$ HiChIP.5kb

Based on APMI, we engineered a new set of features for quantifying the relative contribution of an enhancer **e** to a gene **g** from the gene perspective or enhancer perspective:



In addition, we combined these features to form new features.

fracGmE<sub>eg</sub> = fracGene<sub>eg</sub> \* fracEnh<sub>eg</sub>

 $fracGpE_{eq} = fracGene_{eq} + fracEnh_{eq}$ 



### **E-P** interaction scores are concordant between **EPIC and Enformer**

- Enformer is a deep learning model that predicts gene expression and chromatin states from DNA sequence (Avsec, et al., 2021).
- Enformer contribution scores for hepatocyte enhancers are consistent with EPIC scores on an overall and gene-by-gene basis.







Five-fold cross-validation

Genetic algorithm for feature selection

**□** EPIC enables accurate cell-type-specific prediction of functional E-P interactions using epigenomic data. EPIC outperforms an established method in predicting E-P interactions and in linking GWAS loci to causal genes in a new cell type. Applying EPIC to diverse human cell types may help discover disease-causing genes and enable development of novel therapeutics that target enhancers of disease-related genes.